



Calibration of the Injection Volume for Microinjection of *Xenopus* Oocytes and Embryos

Hazel L. Sive, Robert M. Grainger, and Richard M. Harland

Adapted from [Early Development of *Xenopus laevis*](#), by Hazel L. Sive, Robert M. Grainger, and Richard M. Harland. CSHL Press, Cold Spring Harbor, NY, USA, 2000.

INTRODUCTION

Microinjection of *Xenopus* oocytes or embryos with messenger RNA (mRNA) or DNA is a powerful technique for studying development. Before microinjection can be performed, the injection volume must be calibrated carefully. This protocol describes the calibration procedure for a pressure injector.

RELATED INFORMATION

A protocol is available for [Microinjection of *Xenopus* Oocytes](#) (Sive et al. 2010a). Information is also available on [Microinjection of *Xenopus* Embryos](#) (Sive et al. 2010b).

MATERIALS

Reagents

Paraffin oil

Solution for backfilling the injection needle

Equipment

Injection needles (drawn-out micropipettes)

Microinjector

Microscope, dissecting, with eyepiece micrometer

Microscope slides

Pipettor, automatic, with a long, narrow tip (e.g. the type of tip used to load sequencing gels)

METHOD

1. Backfill the needle using the automatic pipettor and mount on the microinjector.
2. Break the injection needle tip to produce an orifice of $\sim 10\ \mu\text{m}$.
3. Place a small drop of paraffin oil on a microscope slide. Mount the slide on the stage of a dissecting microscope.
4. Calibrate the eyepiece micrometer for the appropriate magnification. Perform a trial injection into the drop of oil.
The injected liquid forms a sphere within the oil droplet.
5. Measure the diameter of the sphere using the eyepiece micrometer. Calculate the injected volume by using the equation $v = 4/3 (\pi r^3)$, where v is the volume and r is the radius of the sphere.
6. Alternative methods for calibrating the injection volume include the following:
 - i. Deposit the drop directly onto the stage micrometer.
The micrometer must be siliconized for the drop to be near spherical.
 - ii. Allow the drop to hang at the end of the needle, where its diameter can be measured using the eyepiece micrometer.
Measure the drop quickly, because evaporation causes it to shrink rapidly.
 - iii. Inject the drop into a dish of paraffin oil. Approximate the volume of the drop using the equation in Step 5.
Alternatively, obtain the volume using the figures in [Table 1](#).
7. Once the appropriate pressure has been established, use the duration of the pressure burst to control the volume precisely.

REFERENCES

1. Sive HL, Grainger RM, Harland RM. 2010a. Microinjection of *Xenopus* oocytes. *Cold Spring Harb Protoc* doi: 10.1101/pdb.prot5536. [[Abstract/Free Full Text](#)]
2. Sive HL, Grainger RM, Harland RM. 2010b. Microinjection of *Xenopus* embryos. *Cold Spring Harb Protoc* doi: 10.1101/pdb.ip81. [[Abstract/Free Full Text](#)]