

**Day 01.**

**Prepare the desire plasmid DNA in a good concentration (Means min conc 0.1 ug/ul) there should not be RNA contamination in preparation.**

**Prepare the Insert DNA ( PCR Product) Clean the product with PCR clean up kit If it is single band product or Gel purify it if it has some nonspecific amplicons.**

**Now set up reaction as follows**

**Restriction Digestion. (for single enzyme)**

**Master mix for Digestion.**

<b>Double distilled water</b>	<b>16 ul</b>
<b>10X buffer</b>	<b>2 ul</b>
<b>Enzyme</b>	<b>2 ul</b>
<b>Mix the master mix by vortex and spin down in microfuge.</b>	

**Reaction tubes**

<b>components</b>	<b>Vector DNA</b>		<b>PCR product or Insert DNA</b>
<b>D D water</b>	<b>X ul (75- rest volume)</b>		<b>Y ul ( 75- rest volume)</b>
<b>10X buffer</b>	<b>7.5ul</b>		<b>7.5ul</b>
<b>DNA</b>	<b>5 ug (Minimum)</b>		<b>All after purification</b>
<b>Enzyme mix</b>	<b>10ul</b>		<b>10ul</b>

**Total Volume==**

**75 ul**

**75 ul**

Incubate at 37 degree for 2 hours and check for digestion with 5ul from Vector DNA tube and 2ul of loading dye on 0.8% Agarose gel.

Lane 1= Marker

Lane 2= Digested DNA ( Vector)

Lane 3 = Undigested Vector DNA.



M 1 2

**Please see the shift in bands.**

**Restriction Digestion. (for two enzyme)**

**Master mix for Digestion.**

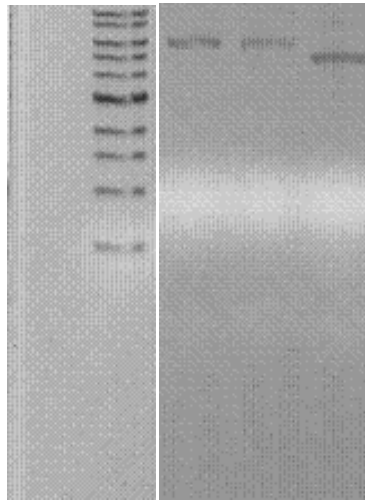
<b>DD Water</b>	<b>16 ul</b>	<b>16 ul</b>
<b>10X buffer</b>	<b>2 ul</b>	<b>2 ul</b>
<b>Enzyme 01</b>	<b>2 ul</b>	<b>00 ul</b>
<b>Enzyme 02</b>	<b>00 ul</b>	<b>2 ul</b>

**Vortex and spin down in a Microfuge.**

**Reaction set up.**

	<b>Vector DNA</b>	<b>Control for Enzyme 01</b>	<b>Control for Enzyme 02</b>	<b>PCR product.</b>
<b>Dd water</b>	<b>75- Rest</b>	<b>20- Rest</b>	<b>20- Rest</b>	<b>75-Rest</b>
<b>10X Buffer</b>	<b>7.5ul</b>	<b>2 ul</b>	<b>2ul</b>	<b>7.5 ul</b>
<b>DNA</b>	<b>Min 5 ug</b>	<b>1 ug plasmid DNA</b>	<b>1 ug Plasmid DNA.</b>	<b>All purified product</b>
<b>Enzyme 01</b>	<b>10 ul</b>	<b>3 ul</b>	<b>00 ul</b>	<b>7 ul</b>
<b>Enzyme 02</b>	<b>10 ul</b>	<b>00 ul</b>	<b>3 ul</b>	<b>7ul</b>
<b>Total Volume</b>	<b>75 ul</b>	<b>20 ul</b>	<b>20 ul</b>	<b>75 ul</b>

**Incubate at 37 Degree for 2 hrs, Check on gel from only control tubes for Enzyme 01 and Enzyme 02**



**M 1 2 3**

**Lane M= Marker , Lane 1 Control tube for Enzyme 01 ,**

**Lane 02 = Control tube for Enzyme 02,**

**Lane 03 = Uncut plasmid.**

**If u are using two restriction enzyme there is no need of treating your digested vector DNA with Alkaline Phosphatase. After digestion is complete u can gel purify the product or let me know about your kit ( I will tell u short cut )**

**But if u r using only one Restriction enzyme treat it with Alkaline Phosphatase Enzyme. See below protocol**

<b>DD Water</b>	<b>100- Rest Volume .</b>
<b>10X Buffer</b>	<b>10 ul</b>
<b>Digested vector DNA</b>	<b>75 ul</b>
<b>Alkaline Phosphatase</b>	<b>2ul</b>
<b>Total volume</b>	<b>100 ul</b>

**Incubate as follows**

**37 Degree= 60 mins ,**

**70 degree = 10 mins**

**4 degree = Pause**

**After treatment is complete u can gel purify the products ( Digested and de phosphorylated Vector DNA and Digested Insert DNA) or let me know about your kit ( I will tell u short cuts also . )**

**Now run Purified Double digested dephosphorylated Vector DNA and digested Insert DNA (Use only 1 ul with DNA Loading dye)**

**Now directly see it on gel with naked eye u shouldsee the intensity of band and approximately analyse how much nano gram DNA is there.**

**( Usually it will be around40-50ng/ul )**

**Now set up the digestion reaction as follows.**

	<b>1:3</b>	<b>1:5</b>	<b>1:7</b>	<b>--Ve</b>	
<b>DDWater</b>					
<b>10X Buffer</b>	<b>1 ul</b>	<b>1 ul</b>	<b>1 ul</b>	<b>1 ul</b>	
<b>DigestedVector DNA</b>	<b>1.5 ul</b>	<b>1.5 ul</b>	<b>1.5 ul</b>	<b>1.5 ul</b>	
<b>Digested Insert DNA</b>	<b>1.2 ul</b>	<b>1.5 ul</b>	<b>2 ul</b>	<b>00 ul</b>	
<b>T4 DNA Ligase enzyme</b>	<b>1 ul</b>	<b>1 ul</b>	<b>1 ul</b>	<b>1 ul</b>	
<b>Total Volume=</b>	<b>10 ul</b>	<b>10 ul</b>	<b>10 ul</b>	<b>10 ul</b>	

**Incubate at 22 degree for 3 hours.**

**Set Bacterial transformation.**