To prepare plasmid DNA by alkaline lysis/PEG treatment:

Note To minimize shearing of contaminating chromosomal DNA, do not use a vortexer during this procedure.

Step Action

1 Pellet 1.5-mL aliquots of culture for 1 minute in a microcentrifuge at maximum speed.

Note A total culture volume of 4.5 mL can be spun down per tube without changing volumes in the procedure. This allows you to achieve a threefold increase in yield while eliminating the need for extra tubes and additional handling.

- 2 Remove the supernatant by aspiration.
- 3 Resuspend the bacterial pellet in 200 µL of GET buffer by pipetting up and down.
- **4** Add 300 μ L of freshly prepared 0.2 N NaOH/1% SDS. Mix the contents of the tube by inversion. Incubate on ice for 5 minutes.
- **5** Neutralize the solution by adding 300 μ L of 3.0 M potassium acetate, pH 4.8. Mix by inverting the tube. Incubate on ice for 5 minutes.
- **6** Remove cellular debris by spinning in a microcentrifuge at maximum speed for 10 minutes at room temperature. Transfer the supernatant to a clean tube.
- **7** Add RNase A (DNase-free) to a final concentration of 20 μ g/mL. Incubate the tube at 37 °C for 20 minutes.
- 8 Extract the supernatant twice with chloroform:
- a. Add 400 µL of chloroform.
- b. Mix the layers by inversion for 30 seconds.
- c. Centrifuge the tube for 1 minute to separate the phases.
- d. Transfer the upper aqueous phase to a clean tube.
- **9** Add an equal volume of 100% isopropanol. Mix the contents of the tube by inversion
- **10** Spin the tube in a microcentrifuge at maximum speed for 10 minutes at room temperature.
- **11** Remove the isopropanol completely by aspiration.
- 12 Wash the DNA pellet with 500 µL of 70% ethanol. Dry under vacuum for 3 minutes.
- 13 Dissolve the pellet in 32 µL of deionized water.
- 14 Add 8.0 μ L of 4 M NaCl, then 40 μ L of autoclaved 13% PEG 8000.
- 15 Mix thoroughly, then leave the sample on ice for 20 minutes.
- **16** Pellet the plasmid DNA by spinning in a microcentrifuge at maximum speed for 15 minutes at 2–6 °C.
- 17 Carefully remove the supernatant. Rinse the pellet with 500 µL of 70% ethanol.
- 18 Resuspend the pellet in 20 µL of deionized water. Store at -15 to -25 °C.